

## Synthesis of Oligonucleotides Containing 5-(Hydroxymethyl)-2'-deoxyuridine at Defined Sites

Lawrence C. Sowers\*<sup>†</sup> and G. Peter Beardsley<sup>‡</sup>

Division of Pediatrics, City of Hope National Medical Center, 1500 East Duarte Road, Duarte, California 91010, and Departments of Pediatrics and Pharmacology, Yale University School of Medicine, 333 Cedar Street, New Haven, Connecticut 06510

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**Summary:** A method is described for the solid-phase synthesis of oligonucleotides containing the DNA oxidation damage product, 5-(hydroxymethyl)-2'-deoxyuridine (HMdU) at selected sites using a phosphoramidite synthon regioselectively protected on the 5-(hydroxymethyl) group.

5-(Hydroxymethyl)-2'-deoxyuridine (HMdU) is a damaged nucleoside formed by hydroxyl radical attack on thymidine residues in DNA. HMdU has been identified in the DNA of cells exposed to oxidizing chemicals and radiation.<sup>1-4</sup> When administered to cells in culture, HMdU is both mutagenic<sup>5,6</sup> and cytotoxic,<sup>7,8</sup> presumably due in part to incorporation into cellular DNA. Attempts to further investigate the deleterious effects of HMdU in DNA using biochemical and physical methods have been hampered by the absence of suitable methods for the preparation of HMdU-containing oligonucleotides. We report here an efficient method for the solid-phase synthesis of oligonucleotides containing HMdU residues at selected sites.

Selective tritylation<sup>9</sup> of the 5'-hydroxyl of HMdU was deemed problematic because the 5- and 5'-(hydroxymethyl) groups are of similar nucleophilicity. Previously, Prusoff and co-workers<sup>10</sup> reacted HMdU with 1 equiv of *p*-toluenesulfonyl chloride and obtained the two monosulfonates, the disulfonate and parent compound. The 5'-tosylated derivative was isolated with difficulty in 12% yield. When following this scheme, however, one is still left with the difficult task of selectively generating base-labile protection for the 5-(hydroxymethyl) group and subsequent preparation of the 3' phosphoramidite.

Recently, Levy and Teebor<sup>11</sup> described the preparation of oligodeoxynucleotides containing HMdU using DNA polymerase and the triphosphate derivative of HMdU. These investigators utilized a biosynthetic approach after encountering severe difficulties during attempts to synthesize a suitably protected phosphoramidite derivative. It was reported that the triphosphate of HMdU is prepared

in only 1-4% yield.<sup>11</sup> Furthermore, the polymerase method<sup>11,12</sup> would be inadequate for the preparation of the quantities of oligonucleotides containing HMdU at selected sites that would be required for physical studies.

Alternatively, it is known that 5-(hydroxymethyl)uracil and derivatives will condense with both alcohols and carboxylic acids in the presence of protic acids.<sup>13,14</sup> Scheit<sup>14</sup> proposed the presence of a resonance-stabilized methylene cation to account for the difference in reactivity of the 5'- and 5-(hydroxymethyl) groups of 5-(hydroxymethyl) nucleosides under acidic conditions. We sought to exploit this selective reactivity to acetylate the 5-(hydroxymethyl) group of 5-(hydroxymethyl)-2'-deoxyuridine (HMdU). HMdU 1 (Scheme I), prepared by reaction of deoxyuridine and paraformaldehyde in aqueous KOH,<sup>10</sup> was refluxed for 30 min in acetic acid containing HCl. While significant conversion occurred, glycosidic bond cleavage was also observed. Trifluoroacetic acid was substituted for the HCl, and the reaction was repeated.<sup>15</sup> Analysis of the reaction components by silica gel thin-layer chromatography indicated the conversion of HMdU to a single product with little glycosidic bond cleavage. Under identical conditions, neither deoxyuridine nor thymidine reacted, consistent with the expected selectivity of the reaction.

Following the reaction of HMdU in acetic acid/trifluoroacetic acid, solvent was evaporated under reduced pressure. The product 2 was isolated as a white solid following silica gel chromatography in 84% yield. Mass spectral analysis of the product 2 indicated that the derivative was monoacetylated, with a mass of 300<sup>16</sup> ( $M - 1 = 299$ , negative ion mode FAB). The composition of 2 was confirmed by elemental analysis. Upon aminolysis (60 °C, 12 h), the acetylated derivative 2 quantitatively regenerated the parent compound, HMdU 1.

The proton NMR spectrum of 2 confirms the site of acetylation. Previously, Frenkel et al.<sup>2</sup> compared the proton NMR spectra of HMdU and the triacetylated derivative formed by reaction of HMdU with acetic anhydride in pyridine. The 5, 5', and 3' protons are well resolved for both HMdU and the triacetylated derivative. It was shown that acetylation shifts proton resonances for the 5, 5', and 3' protons downfield by 0.51, 0.52, and 0.87 ppm, respectively. For the monoacetylated derivative 2, only the 5-methylene proton resonance was observed to shift downfield significantly (0.49 ppm) relative to parent compound 1.

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(15) HMdU 1 was selectively acetylated on the 5-(hydroxymethyl) group by reflux in acetic acid containing catalytic trifluoroacetic acid. The acetylated derivative was isolated by silica gel chromatography in 84% yield.

(16) The composition of 2 was verified by mass spectrometry, elemental analysis, and proton NMR spectroscopy.

\* Author to whom correspondence should be addressed.

<sup>†</sup> City of Hope National Medical Center.

<sup>‡</sup> Yale University School of Medicine.

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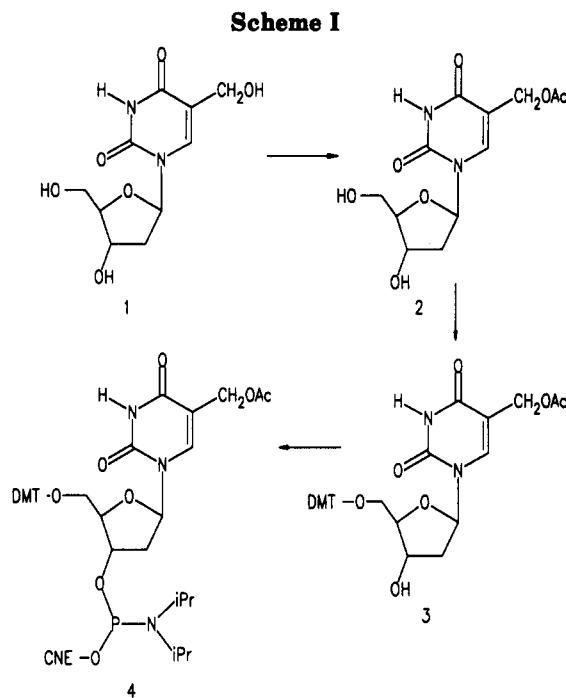
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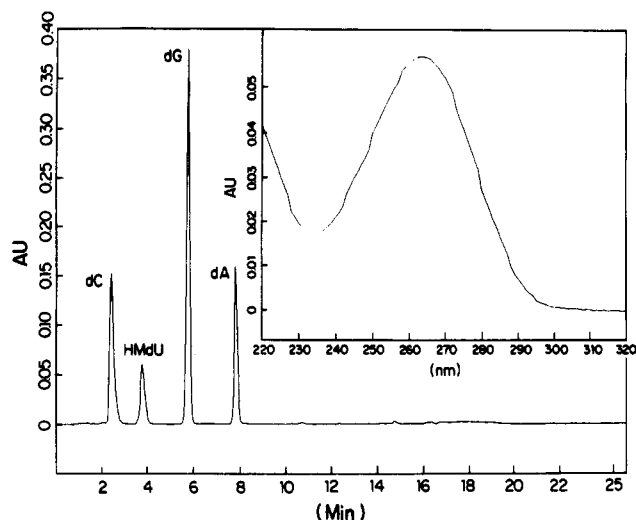
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Once acetylated, the HMdU derivative can be treated essentially as thymidine. The 5'-*O*-(4,4'-dimethoxytrityl) **3** and 3'-*O*-(2-cyanoethyl)-*N,N*-diisopropylphosphoramidite **4** derivatives were prepared by standard methods.<sup>9</sup> A seven-base oligodeoxynucleotide of sequence 3'd(CGGHGAC) was then prepared by manual solid-phase methods at the 15  $\mu$ mol scale. All coupling yields, including that with the HMdU synthon, were >98% as indicated by release of trityl cation. Following the synthesis, the oligo was deprotected in aqueous ammonia at 60 °C, 12 h. The tritylated oligo was first purified by HPLC with a PRP column, followed by purification of the detritylated oligo by reversed-phase HPLC eluted with aqueous triethylammonium acetate and acetonitrile.

The purified seven-base oligo was homogeneous by HPLC. A fraction was enzymatically hydrolyzed with nuclease P1, followed by bacterial alkaline phosphatase.<sup>17</sup> The oligo was digested to completion, indicating the correct formation of 3'-5' internucleotide phosphate linkages. The resulting deoxynucleosides were analyzed by reversed-phase HPLC. As shown in Figure 1, peaks corresponding to the four bases present in the oligo were observed in the correct ratios based upon integration of the chromatogram at 260 nm (Figure 1). The UV spectrum of the HMdU peak eluting at 3.72 min (Figure 1, inset) was consistent with published data.<sup>18</sup> Neither the acetylated derivative nor unknown peaks were observed.

In this context, the quantitation of HMdU in DNA exposed to oxidizing conditions has been established by



**Figure 1.** HPLC analysis of deoxynucleosides derived from the enzymatic hydrolysis of a seven base oligonucleotide 3'd-(CGGHGAC) containing HMdU. The labels indicate the identity of the peaks as determined by coincident retention times with authentic samples. Inset: UV spectrum of the HMdU peak (3.72 min); abs max, 264 nm; abs min 234 nm.

HPLC methods<sup>19</sup> since GC/MS methods were unsatisfactory for this purpose.<sup>20</sup> Deoxynucleosides for HPLC analysis are generated by enzymatic hydrolysis. However, for the GC/MS method, DNA is hydrolyzed with formic acid prior to trimethylsilylation. Reaction of HMdU under acidic conditions, which has been exploited here for the selective protection of HMdU, may likely explain the failure to detect (hydroxymethyl)uracil by the GC/MS method. The reactivity of HMdU derivatives under acidic conditions further suggests possible strategies for the selective derivatization of HMdU prior to either HPLC or GC/MS analysis.

The synthesis reported here provides a facile method for preparing oligonucleotides containing HMdU at specific positions suitable for biochemical and physical studies. This protection strategy also provides a method for generating additional nucleoside analogues and oligonucleotides containing modifications to the 5-position, including the radiolabeling of derivatives for quantitative analysis.

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**Supplementary Material Available:** Experimental procedures (5 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

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